

See discussions, stats, and author profiles for this publication at:
<https://www.researchgate.net/publication/7775031>

Essential oils of commonly used plants as inhibitors of peroxynitrite-induced tyrosine nitration

Article in *Fitoterapia* · August 2005

DOI: 10.1016/j.fitote.2005.04.010 · Source: PubMed

CITATIONS

8

READS

36

4 authors, including:



Silvio Chericoni

Azienda Ospedaliero-Universitaria Pisana

34 PUBLICATIONS 687 CITATIONS

[SEE PROFILE](#)



Jose Maria Prieto

University College London

89 PUBLICATIONS 1,596 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



Prostate Cancer [View project](#)



New Analytical Techniques for Herbal Medicines Quality Control [View project](#)

All content following this page was uploaded by [Silvio Chericoni](#) on 26 November 2015.

The user has requested enhancement of the downloaded file. All in-text references [underlined in blue](#) are added to the original document and are linked to publications on ResearchGate, letting you access and read them immediately.



Short report

Essential oils of commonly used plants as inhibitors of peroxyxynitrite-induced tyrosine nitration

Silvio Chericoni *, Josè M. Prieto, Patrizia Iacopini, Ivano Morelli

Dipartimento di Chimica Bioorganica e Biofarmacia, Università di Pisa, via Bonanno 33, 56126 Pisa, Italy

Received 20 January 2005; accepted 26 April 2005

Abstract

The essential oils obtained from fifteen relevant and commonly used plants belonging to Cruciferae, Lamiaceae, Lauraceae, Apiaceae, and Zingiberaceae were screened using an in vitro model of peroxyxynitrite-induced tyrosine nitration. Almost complete inhibition of 3-nitrotyrosine formation (91% at 300 µg/ml) was achieved only with the essential oil obtained from the leaves of *Laurus nobilis*. 1,8-Cineol, accounting for a 50% of this essential oil, which resulted as inactive in this model, thus evidencing a major role for the minor volatile compounds present in the leaves.

© 2005 Elsevier B.V. All rights reserved.

Keywords: *Laurus nobilis*; Peroxyxynitrite; 3-Nitrotyrosine; Essential oil; 1,8-Cineol

1. Plant

Botanical name, family, and investigated part of the plants are reported in [Table 1](#). All the plant material was purchased from the local market except *Sinapis alba*, *Zingiber*

* Corresponding author.

E-mail address: chersil@farm.unipi.it (S. Chericoni).

Table 1
In vitro inhibition of 3-nitrotyrosine formation by selected plants' essential oils

Plants	Common name	Part used	%I ^a
Cruciferae			
<i>Sinapis alba</i>	Mustard	Fruits	0
Lamiaceae			
<i>Lavandula spica</i> × <i>latifolia</i>	Spike lavender	Flowers	0
<i>Lavandula officinalis</i>	Lavender	Flowers	0
<i>Origanum majorana</i>	Oregano	Aerial parts	8
<i>Rosmarinus officinalis</i>	Rosemary	Aerial parts	21
<i>Salvia officinalis</i>	Sage	Aerial parts	0
<i>Thymus vulgaris</i>	Thyme	Aerial parts	36
<i>Mentha spicata</i>	Mint	Aerial parts	38
Lauraceae			
<i>Laurus nobilis</i>	Bay	Leaves	91
Apiaceae			
<i>Carumi carvi</i>	Caraway	Fruits	0
<i>Coriandrum sativum</i>	Coriander	Fruits	45
<i>Foeniculum vulgare</i>	Sweet fennel	Fruits	39
<i>Pimpinella anisum</i>	Anise	Fruits	0
Zingiberaceae			
<i>Zingiber officinalis</i>	Ginger	Roots	0
<i>Curcuma zedoaria</i>	Zedoary	Roots	0
1,8-Cineol (500 µM)	–	–	0
Ascorbic acid (405 µM)	–	–	50

^a Percentage of in vitro inhibition of 3-nitrotyrosine formation (essential oils were assayed at a final concentration of 300 µg/ml).

officinalis, *Curcuma zedoaria*, *Carumi carvi*, and *Coriandrum sativum* that were obtained from RES Pharma s.r.l. (Milano, Italy).

2. Uses in traditional medicine

Antimicrobial, antifungal, digestives, mouthwashes, wound healing, and balsamic actions [1].

3. Previously isolated classes of constituents

Terpenoids [1].

4. Tested material

Essential oils were obtained by hydrodistillation in a Clevenger apparatus for 2 h as described in the Italian Pharmacopea XI ed. 1,8-Cineol and ascorbic acid were purchased from Sigma-Aldrich.

5. Studied activity

Inhibition of in vitro peroxyxynitrite-induced tyrosine nitration. Briefly, the reaction was carried out adding, under vigorous vortexing, the peroxyxynitrite synthesised according Beckman et al. [2] (1 mM final concentration) to a solution containing the essential oil or pure compounds at the desired concentrations, tyrosine (2 mM) and HCO_3^- (50 mM) all dissolved in 50 mM phosphate buffer (pH 7.4). Test compounds were dissolved in MeOH (final concentration of 0.5%) and kept into ice till used. Controls, with or without MeOH, were always performed to detect any interference of the solvent with the tests. Quantitative determination of the formed 3-nitrotyrosine was performed by HPLC–UV using an external standard calibration curve ($r^2=0.999$). Ascorbic acid was used as a positive reference compound.

6. Results

The composition of the essential oil obtained from leaves of *L. nobilis* was determined by GC–MS and resulted to be: 1,8 cineol (50%), sabinene (9%), linalool and α -terpinyl-acetate (6% each), α -pinene (5%), β -pinene and methyleugenol (4% each), and other identified minor compounds (2–0.5%) accounting for 98.84% of the total oil composition. Complete results are reported in Table 1.

7. Conclusions

This is the first report on the activity of essential oils as inhibitors of peroxyxynitrite mediated processes. Nitration of tyrosine residues in key proteins is a process mediating many acute and chronic pathologies [3] and 3-nitrotyrosine is considered a biomarker of the oxidative stress [4]. In this screening, the essential oil obtained from leaves of *L. nobilis* provided the highest in vitro protection against tyrosine nitration. Its main component, 1,8-cineol, resulted inactive, evidencing that minor compounds and/or complex interactions between the components of the essential oil play an important role.

References

- [1] Bruneton J. Pharmacognosie, Phytochimie, Plantes Medicinales, 3rd ed. Paris: Tec et Doc; 1999.
- [2] Beckman JS, Chen J, Ischiropoulos H, Crow JP. Methods Enzymol 1994;233:229.
- [3] Virág L, Szabó É., Gergely P, Szabó C. Toxicol Lett 2003;140:113.
- [4] Althaus JS, Schmidt KR, Fountain S, Tseng MT, Carroll TR, Galatsis P, et al. Free Radic Biol Med 2000;29:1085.